

O₂ and CO Binding to a New Type of Iron(II) Porphyrins

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In hemoproteins, the discrimination against the binding of CO relative to that of O₂ plays an important role in preventing poisoning by CO. Distal effects [1] functionally produced by globin are thought to be a major motive force for this discrimination and can be separated into two effects: polar and steric. To elucidate the role of these effects on the discrimination, studies [2–5] have been carried out using wide varieties of model porphyrins on the effects upon the discrimination. Nevertheless, whether so-called ‘cavity’ within globin discriminates between O₂ and CO by a steric effect is still unresolved [2a, 4b]. To clarify further the steric effect, a series of model porphyrins were designed and synthesized with the unique structure in which one aliphatic chain crosses over another aliphatic chain bridging the same side of a porphyrin plane (Fig. 1).

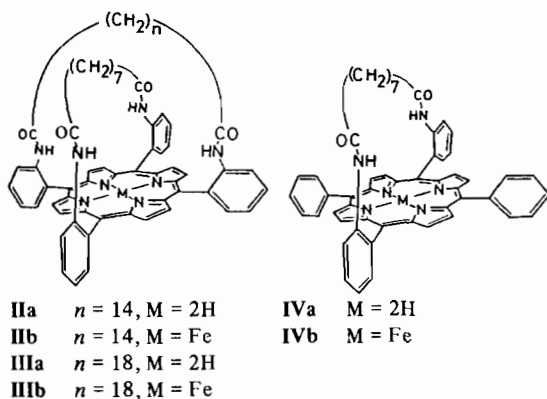


Fig. 1. Model porphyrins.

The discrimination between O₂/CO binding by steric effect was studied experimentally on these model porphyrins.

5 α ,15 α -Bis(2-aminophenyl)-10 α ,20 α -(2,2'-nonanediamidodiphenyl)porphyrin, (**I**) was prepared from the isomerization of 2-aminophenyl groups of its 5 β ,15 β -isomer [6]. Porphyrins **IIa** and **IIIa** were synthesized from coupling **I** with (CH₂)_n(COCl)₂ ($n = 14$ or 18) under high dilution conditions [6, 7]. Porphyrin **IVa** was prepared by the reported method [8]. All new compounds described gave spectral (FAB mass, ¹H NMR) and analytical data in accor-

dance with their assigned structures. It is expected that the -(CH₂)₁₄- or -(CH₂)₁₈- chain in **IIa** or **IIIa** gives steric repulsion to the heptane chain and makes the chain close to the porphyrin ring. Also the heptane chain of **IIa** is expected to receive more severe steric repulsion than that of **IIIa**, because the crossing over chain of **IIa** (*i.e.* the -(CH₂)₁₄- chain) is shorter than the -(CH₂)₁₈- chain of **IIIa**. This expectation was confirmed by ¹H NMR measurements. As shown in Table I, the methylene proton signals at the 4-position in the heptane chain of **IIa** and **IIIa** shift upfield by 1.24 and 0.62 ppm, respectively, compared to that of **IVa**. These upfield shifts are the result of the ring current of porphyrin [9], so the heptane chain of **IIa** is suggested to be more crushed than that of **IIIa**. Iron insertions into **IIa** or **IIIa** and **IVa** were accomplished by heating **IIa** or **IIIa** in acetic acid and **IVa** in tetrahydrofuran, respectively, with FeBr₂. Purifications of iron(III) porphyrins were carried out by silica-gel column chromatography. Fe(III) complexes were reduced to Fe(II) complexes by aqueous sodium dithionite in a two-phase system [3a] (toluene–water) under Ar in the presence of 1,2-dimethylimidazole (1,2-Me₂Im). The Fe(II) complexes prepared in this study bind O₂ or CO reversibly in toluene at 20 °C in the presence of 1,2-Me₂Im and their half-saturation pressures for O₂ or CO binding were obtained by spectroscopic titrations [6] (Fig. 2 and Table II).

The O₂ affinity of **IIIb** is slightly higher but the CO affinity is 2-fold lower than that of **IVb**. Also the O₂ and CO affinities of **IIb** are about 100 and 600 times lower, respectively, than those of **IIIb**. That is, **IIb** reduces CO affinity selectively and has a remarkably low value of $M = 24$. Recently, Traylor *et al.* [4b] reported the model complex 3,5-pyridine-5,5-hemecyclophane, which has a small M value. They proposed that a polar effect induced by pyridine is the principal factor in the discrimination. The changes in O₂ and CO affinities between **IIb** and **IVb** can be explained by the difference in local polarity. That is, the cavity of **IIIb** must be more

TABLE I. ¹H NMR Data^a for Nonanediamido Groups in Porphyrins

Porphyrins	Methylene protons ^b				Amido protons
	1,7-	2,6-	3,5-	4-	
IVa	1.16	-0.51	-1.24	-2.51	5.99
IIIa	1.25	-1.16	-1.51	-3.13	6.31
IIa	1.21	-1.58	-1.58	-3.75	6.45

^aChemical shift (ppm) from TMS in CDCl₃.

^bNumbers refer to positions in the heptane group.

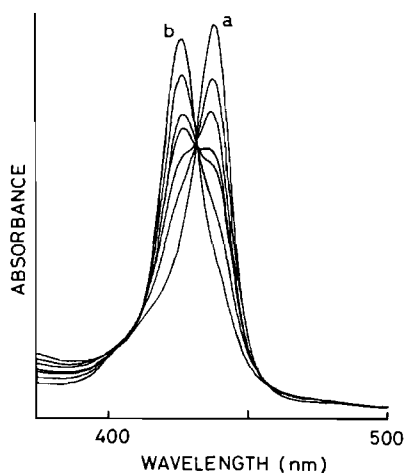


Fig. 2. Determination of $P_{1/2}$ (CO) for **IIb**: 0.5 M 1,2-dimethylimidazole in toluene at 20 °C: (curve a) under 1 atom of N_2 ; (curve b) CO partial pressure of 395 torr. The following partial pressures of CO were used: 16.0, 31.4, 60.3, 81.2 and 161 torr.

TABLE II. O_2 and CO Binding to Iron(II) Complexes

Complexes ^a	$P_{1/2}$ (O_2) ^b (torr)	$P_{1/2}$ (CO) ^b (torr)	M^c
IVb	18	0.05	360
IIIb	15	0.09	170
IIb	1400	62	23

^aSolutions are 0.5 M 1,2-dimethylimidazole in toluene at 20 °C. ^b $P_{1/2}$ (O_2) and $P_{1/2}$ (CO) are half-saturation pressures for O_2 and CO binding, respectively. ^c $M = P_{1/2}$ (O_2)/ $P_{1/2}$ (CO).

polar than that of **IVb**, because **IIIb** has two more amido groups in the cavity than **IVb** has. On the contrary, the discrimination reaction observed in **IIb** may be responsible for steric hindrance to bound O_2 and CO, respectively, for the following reasons. The change in the polarity within cavities between

IIb and **IIIb** is small compared to that between **IIIb** and **IVb**, since both **IIb** and **IIIb** have four similar amido groups. On the other hand, 1H NMR results for free base porphyrins imply that the strapped heptane chain is crushed in the order **IVb** (without crushing) < **IIIb** < **IIb**, and the steric hindrance to bound O_2 and CO will be increased in the same order. In particular, the Fe–CO bond unit prefers to be linear and normal to the porphyrin plane in Fe(II)–porphyrin systems, whereas O_2 binds in a bent fashion [2]. Therefore the Fe–CO unit will receive more severe steric hindrance than that of the Fe– O_2 one. Thus, it is concluded that steric hindrance induced by the crushed heptane chain in **IIb** causes a remarkable reduction in CO affinity but a minor one in O_2 affinity, respectively.

References

- (a) A. Szabo, *Proc. Natl. Acad. Sci. U.S.A.*, **75**, 2108 (1978); (b) K. Moffat, J. F. Deatherage and D. W. Seybert, *Science (Washington, D.C.)*, **206**, 1035 (1979); (c) J. M. Baldwin, *J. Mol. Biol.*, **136**, 103 (1980).
- (a) J. P. Collman, J. I. Brauman, L. I. Brent, J. L. Sessler, R. M. Morris and Q. H. Gibson, *J. Am. Chem. Soc.*, **105**, 3052 (1983); (b) J. P. Collman, J. I. Brauman and K. M. Doxsee, *Proc. Natl. Acad. Sci. U.S.A.*, **76**, 6035 (1979).
- (a) T. Hashimoto, R. L. Dyer, M. J. Crossley, J. E. Baldwin and F. Basolo, *J. Am. Chem. Soc.*, **104**, 2101 (1982); (b) M. Shimizu, F. Basolo, M. N. Vallejo and J. E. Baldwin, *Inorg. Chim. Acta*, **91**, 247 (1984).
- (a) T. G. Traylor, S. Tsuchiya, D. Campbell, M. Mitchell, D. Stynes and N. Koga, *J. Am. Chem. Soc.*, **107**, 604 (1985); (b) T. G. Traylor, N. Koga and L. A. Deardurff, *J. Am. Chem. Soc.*, **107**, 6504 (1985).
- D. Lavalette, C. Tetreau, J. Mispelter, M. Momenteau and J. M. Lhoste, *Eur. J. Biochem.*, **145**, 555 (1984).
- Y. Uemori, A. Nakatsubo, H. Imai, S. Nakagawa and E. Kyuno, *Inorg. Chim. Acta*, **124**, 153 (1985).
- J. P. Collman, J. I. Brauman, T. J. Collins, B. L. Iverson, G. Lang, R. B. Pettman, J. L. Sessler and M. A. Walters, *J. Am. Chem. Soc.*, **105**, 3038 (1983).
- Y. Uemori and E. Kyuno, *Inorg. Chim. Acta*, **125**, L45 (1986).
- R. J. Abraham, S. C. M. Fell and K. M. Smith, *Org. Magn. Reson.*, **9**, 367 (1977).